

BRIEF REPORT

The Uridine Diphosphate Glucuronosyltransferase 2B15 D⁸⁵Y and 2B17 Deletion Polymorphisms Predict the Glucuronidation Pattern of Androgens and Fat Mass in Men

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Context: Previous *in vitro* studies have demonstrated that the UDP glucuronosyltransferase (UGT)2B15 and UGT2B17 glucuronidate androgens and their metabolites.

Objective: Our objective was to determine *in vivo* whether the UGT2B15 D⁸⁵Y and the UGT2B17 deletion polymorphisms predict androgen glucuronidation and body composition.

Participants: Two population-based cohorts including young adult (n = 1068; age = 18.9 yr) and elderly (n = 1001; age = 75.3 yr) men were included in the study.

Main Outcome Measures: Serum and urine levels of testosterone (T) and dihydrotestosterone (DHT) were measured by gas chromatography-mass spectrometry, and serum levels of the major glucuronidated androgen metabolites androstane-3 α ,17 β -diol (androstenediol)-3-glucuronide, androstenediol-17-glucuronide, and androsterone-glucuronide were measured by liquid chromatography-tandem mass spectrometry. Body composition was measured by dual-energy x-ray absorptiometry.

Results: Both the UGT2B15 D⁸⁵Y and the UGT2B17 deletion polymorphisms were associated with serum levels of androstan-

ediol-17-glucuronide ($P < 0.001$) but not with levels of androstenediol-3-glucuronide or androsterone-glucuronide in both cohorts. Glucuronidation of T and DHT was associated with the UGT2B17 deletion but not with the UGT2B15 D⁸⁵Y polymorphism, suggested by strong associations between the deletion polymorphism and urine levels of these two hormones. Both polymorphisms were associated with several different measures of fat mass ($P < 0.01$). The UGT2B17 deletion polymorphism was associated with insulin sensitivity ($P < 0.05$) as indicated by the homeostasis model assessment index.

Conclusions: The UGT2B15 D⁸⁵Y and the UGT2B17 deletion polymorphisms are both predictors of the glucuronidation pattern of androgens/androgen metabolites. Our findings indicate that UGT2B17 is involved in 17-glucuronidation of mainly T but also of DHT and androstenediol and that UGT2B15 is involved in the 17-glucuronidation of androstenediol. Furthermore, these two polymorphisms are predictors of fat mass in men. (*J Clin Endocrinol Metab* 92: 4878–4882, 2007)

CONJUGATION OF ANDROGENS with glucuronic acid has been suggested to play a role in the regulation of the intracellular levels of unconjugated androgens as well as their biological activities in tissues (1, 2).

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Abbreviations: CV, Coefficient of variation; Del, deletion; DHT, dihydrotestosterone; DXA, dual-energy x-ray absorptiometry; HOMA, homeostasis model assessment; T, testosterone; UGT, UDP glucuronosyltransferase; WT, wild type.

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Circulating levels of androstane-3 α ,17 β -diol (androstenediol)-glucuronides and androsterone-glucuronide, the two major 5 α -reduced androgen metabolites in serum in men, originate from both the testicular androgen testosterone (T) and the adrenal C₁₉ steroids (3). Androstenediol can be glucuronidated either at the 3 α -hydroxy position (androstenediol-3-glucuronide) or at the 17 β -hydroxy position (androstenediol-17-glucuronide). UDP glucuronosyltransferase (UGT)2B15 and UGT2B17 are two important enzymes for the glucuronidation of androgens and their metabolites (2). There is a G to T polymorphism in the UGT2B15 gene, resulting in an aspartate (D⁸⁵) to tyrosine

(Y⁸⁵) amino acid change at position 85 (4). A 150-kb deletion polymorphism spanning the whole *UGT2B17* gene has been identified (5, 6).

The aim of the present study was to determine *in vivo* whether the D⁸⁵Y polymorphism of the *UGT2B15* gene and/or the deletion polymorphism of the *UGT2B17* gene predict the glucuronidation pattern of androgens/androgen metabolites and body composition in men.

Subjects and Methods

Study subjects

Young adult men from the population-based Gothenburg Osteoporosis and Obesity Determinants (GOOD) study ($n = 1068$, 18.9 ± 0.5 yr of age) (7) and elderly men from the Gothenburg part of the population-based MrOS Sweden cohort ($n = 1001$, 75.3 ± 3.2 yr of age) (8) were included. Information about prevalent diabetes mellitus ($n = 110$) was obtained through questionnaires in the elderly cohort. Informed consent was obtained from all study participants.

Dual-energy x-ray absorptiometry (DXA)

Body composition was assessed using Lunar Prodigy DXA for the young adult men (GE Lunar Corp., Madison, WI) or Hologic QDR 4500/A-Delphi for the elderly men (Hologic, Waltham, MA).

Assessment of sex hormones in serum

Serum levels were measured by validated high-sensitive gas chromatography-mass spectrometry [T limit of detection of 0.05 ng/ml and interassay coefficient of variation (CV) of 3.4% dihydrotestosterone (DHT) limit of detection of 0.02 ng/ml and interassay CV of 4.1%] and liquid chromatography-tandem mass spectrometry (androsterone-glucuronide limit of detection of 2.00 ng/ml and interassay CV of 3.7%, androstenediol-3-glucuronide limit of detection of 0.50 ng/ml and interassay CV of 10.7%, androstenediol-17-glucuronide limit of detection of 0.50 ng/ml and interassay CV of 5.3%) as previously described (9, 10). The serum samples in the MrOS cohort were fasting samples obtained before 0900 h. Serum samples in the GOOD study were nonfasting samples obtained over the whole day.

Assessment of sex hormones in urine

Urinary unconjugated steroids (typically < 1% of glucuronide fraction) including T and DHT plus their glucuronides were determined by gas chromatography-mass spectrometry ($n = 449$, randomized subsample of the Gothenburg part of MrOS) after hydrolysis of the conjugates with β -glucuronidase as previously described (11, 12). Interassay CV was less than 10% for all steroids analyzed. Urine samples were spot collections (before 0900 h).

Serum/plasma levels of SHBG, insulin, and glucose

SHBG was measured using immunoradiometric assay (Orion Diagnostics, Espoo, Finland) with interassay CV of 7%. Fasting serum insulin was measured with an immunometric method based on chemiluminescence technology on an ADVIA Centaur (Bayer AB, Göteborg, Sweden) with interassay CV of less than 10%. Fasting plasma glucose was quantitated by an enzymatic method on a Modular (Roche, Stockholm, Sweden) with an interassay CV of less than 4%. Homeostasis model assessment (HOMA) index was calculated as the product of fasting serum insulin level (micro-units per milliliter) and fasting plasma glucose level (millimoles per liter) divided by 22.5.

Genetic polymorphism analysis

Genotyping of the *UGT2B15* D⁸⁵Y (rs 1902023) single nucleotide polymorphism and the *UGT2B17* deletion polymorphism was performed as previously described (4, 5, 13). Due to massive manual workload [separation on gel for the deletion (Del) detection], the *UGT2B17* deletion polymorphism was analyzed in only a subsample ($n = 615$, subjects with DNA available and starting from the last recruited subject) of the young adult male cohort; only those not found in the assay detecting the intact *UGT2B17* gene [wild-type (WT)] were chosen for *UGT2B17* homozygous Del allele identification.

Statistical analysis

Existence of Hardy-Weinberg equilibrium was tested using χ^2 analysis. Linkage disequilibrium between the two polymorphisms was calculated by a likelihood ratio test using the genetic analysis package (version 1.0-9) and LD function of the R 2.4.0 program (www.r-project.org). Differences in serum and urinary sex steroid and metabolite levels between the different genotypes (DD, DY, and YY; and Del/Del, Del/WT, and WT/WT) were compared by ANOVA, followed by Tukey's *post hoc* test. The independent predictive value of the *UGT2B15* D⁸⁵Y and the *UGT2B17* deletion polymorphisms for parameters reflecting body composition and serum parameters were investigated using linear regression analyses including age as covariate.

Results

Allele frequencies for D and Y in the *UGT2B15* D⁸⁵Y polymorphism were 0.47 and 0.53, respectively, in the young adult men and 0.45 and 0.55, respectively, in the elderly men. For the *UGT2B17* deletion polymorphism, allele frequencies were 0.31 for the deletion and 0.69 for the WT allele in the elderly men. Neither in the young adult nor in the elderly men did χ^2 analyses show any deviation from Hardy-Weinberg equilibrium for any of the polymorphisms. The results from the linkage disequilibrium analysis showed no evi-

TABLE 1. The *UGT2B17* deletion and the *UGT2B15* D⁸⁵Y polymorphisms as predictors of sex steroids and SHBG in elderly men

	All subjects	<i>UGT2B17</i> deletion				<i>UGT2B15</i> D ⁸⁵ Y			
		Del/Del	Del/WT	WT/WT	<i>P</i> value	DD	DY	YY	<i>P</i> value
Serum parameters		(n = 77)	(n = 432)	(n = 444)		(n = 213)	(n = 492)	(n = 271)	
T (ng/ml)	4.43 ± 1.87 ^a	4.07 ± 1.84	4.37 ± 1.82	4.55 ± 1.91	NS	4.64 ± 2.00	4.42 ± 1.92	4.31 ± 1.69	NS
DHT (ng/ml)	0.35 ± 0.19 ^a	0.32 ± 0.17	0.36 ± 0.21	0.36 ± 0.19	NS	0.37 ± 0.21	0.35 ± 0.20	0.35 ± 0.17	NS
Androstosterone-G (ng/ml)	33.3 ± 67.1 ^a	26.8 ± 19.5	36.2 ± 98.2	31.1 ± 24.0	NS	33.1 ± 29.0	33.3 ± 90.4	32.2 ± 31.7	NS
Androstenediol-3G (ng/ml)	1.50 ± 2.24	1.31 ± 0.88	1.61 ± 3.18	1.40 ± 1.02	NS	1.51 ± 1.17	1.47 ± 2.89	1.48 ± 1.44	NS
Androstenediol-17G (ng/ml)	2.64 ± 2.47 ^a	1.64 ± 1.27 ^c	2.86 ± 3.16 ^b	2.61 ± 1.76 ^b	<0.001	3.20 ± 2.03 ^{b,c}	2.64 ± 2.59 ^b	2.15 ± 2.48 ^c	<0.001
SHBG (nmol/liter)	47.3 ± 24.7 ^a	44.5 ± 18.4	49.1 ± 26.3	46.0 ± 23.9	NS	45.8 ± 22.8	48.3 ± 25.9	46.8 ± 23.4	NS
Urine parameters		(n = 34)	(n = 227)	(n = 181)		(n = 88)	(n = 227)	(n = 134)	
T (ng/μmol creatinine)	1.88 ± 1.19	0.15 ± 0.10 ^c	1.89 ± 1.05 ^b	2.22 ± 1.22 ^b	<0.001	1.91 ± 1.09	1.80 ± 1.23	2.03 ± 1.21	NS
T/epitestosterone	1.02 ± 0.76	0.08 ± 0.04 ^c	1.05 ± 0.78 ^b	1.17 ± 0.69 ^b	<0.001	1.05 ± 0.72	0.97 ± 0.82	1.09 ± 0.69	NS
DHT (ng/μmol creatinine)	0.34 ± 0.29	0.20 ± 0.27 ^c	0.35 ± 0.30 ^b	0.35 ± 0.28 ^b	0.015	0.39 ± 0.30	0.34 ± 0.32	0.28 ± 0.21	NS

Values are given as means ± SD. Serum hormone levels were compared by Student's *t* test and *P* values by ANOVA. G, Glucuronide; NS, nonsignificant.

^a *P* < 0.05 vs. young adult men.

^b *P* < 0.05 vs. Del/Del for *UGT2B17* deletion and *P* < 0.05 vs. YY for *UGT2B15* D⁸⁵Y.

^c *P* < 0.05 vs. Del/WT for *UGT2B17* deletion and *P* < 0.05 vs. DY for *UGT2B15* D⁸⁵Y.

dence of linkage between the UGT2B15 D⁸⁵Y and the UGT2B17 deletion polymorphisms.

Serum and urine analyses

Serum levels of the glucuronidated androgen metabolite androstenediol-17-glucuronide, but not those of androstenediol-3-glucuronide or androsterone-glucuronide, were associated with the UGT2B17 deletion polymorphism in both elderly and young adult men (Tables 1 and 2). Androstenediol-17-glucuronide levels were lower for subjects with the Del/Del genotype than for subjects with the Del/WT and the WT/WT genotypes. The UGT2B17 deletion polymorphism was strongly associated with the urinary levels of T and moderately associated with urinary DHT (Table 1). Both urinary T and DHT were lower for the Del/Del subjects than for the Del/WT and WT/WT subjects (Table 1). Furthermore, the urinary T to epitestosterone ratio was strongly associated with the UGT2B17 deletion polymorphism.

Androstenediol-17-glucuronide levels were clearly associated with the UGT2B15 D⁸⁵Y polymorphism in both young adult and elderly men (Tables 1 and 2). The serum levels of androstenediol-17-glucuronide were higher for subjects with the DD and the DY genotypes than for subjects with the YY genotype. Serum DHT and SHBG levels but not T levels were slightly higher for the DD than for the YY genotype in the young adult men (Table 2).

Body composition analyses

Several indicators of body fat (body weight, body mass index, total body fat, and trunk fat; $P < 0.05$) were associated with the UGT2B17 deletion polymorphism in the elderly cohort and with the UGT2B15 D⁸⁵Y polymorphism in the young adult cohort (Table 3). Some of the parameters reflecting fat mass (total body fat percent and arm fat; $P < 0.05$) were associated with the UGT2B15 D⁸⁵Y polymorphism also in the elderly cohort. Subjects with the UGT2B15 YY genotype had a higher amount of fat than the subjects with the DY and DD genotypes, and subjects with the UGT2B17 Del/Del genotype had a higher amount of fat than the subjects with the Del/WT and WT/WT genotypes (Table 3; $P < 0.05$). In addition, the UGT2B17 deletion polymorphism was a predictor of serum insulin and HOMA index in the elderly cohort (Table 3; $P < 0.05$).

Discussion

We here demonstrate that both the UGT2B15 D⁸⁵Y polymorphism and the UGT2B17 deletion polymorphism have an impact on the glucuronidation pattern of androgens/androgen metabolites.

The present *in vivo* data, showing that the UGT2B15 D⁸⁵Y polymorphism is strongly associated with serum levels of androstenediol-17-glucuronide but not androstenediol-3-glucuronide or androsterone-glucuronide (Fig. 1) support earlier *in vitro* findings that UGT2B15 specifically conjugates the 17 β -hydroxy position of androstenediol (14). Furthermore, the results indicate that the G to T polymorphism in the UGT2B15 gene is functional. Because the serum levels of androstenediol-17-glucuronide were higher in the DD than in the YY subjects, we conclude that subjects with the DD genotype probably have a more efficient UGT2B15 enzyme for 17-glucuronidation of androstenediol than the YY subjects.

Earlier *in vitro* studies have indicated that UGT2B17 has the capacity to glucuronidate androstenediol at the 17 β -hydroxy position (resulting in androstenediol-17-glucuronide) and androsterone at the 3 α -hydroxy position (resulting in androsterone-glucuronide) (2). However, the present *in vivo* study of a naturally occurring gene inactivation of the UGT2B17 gene demonstrates that UGT2B17 is a rather selective enzyme in the glucuronidation of androgen metabolites, enhancing the 17 β -glucuronidation of androstenediol but not the 3 α -glucuronidation of androstenediol or androsterone (Fig. 1).

UGTs glucuronidate not only androgen metabolites but also T and DHT. The present finding of a major role of UGT2B17 for T excretion in elderly men (Fig. 1) confirms our previous results in young adult men (13) and is consistent with previous *in vitro* reports showing that T is a good substrate for UGT2B17 (15) but not for UGT2B15 (2). In addition, the present finding that urine DHT was associated with the UGT2B17 deletion polymorphism but not with the UGT2B15 D⁸⁵Y polymorphism (Fig. 1) is supported by previous studies investigating enzymatic activity *in vitro* (2, 14). Interestingly, the clear association between the UGT2B15 D⁸⁵Y polymorphism and androstenediol-17-glucuronide levels was accompanied by slightly affected serum levels of SHBG and DHT in the young adult men, suggesting that this polymorphism might affect androgen-dependent phenotypes. The mechanism behind the affected SHBG levels in the young adult

TABLE 2. The UGT2B17 deletion and the UGT2B15 D⁸⁵Y polymorphisms as predictors of sex steroids and SHBG in young adult men

Serum parameters	All subjects	UGT2B17 deletion			UGT2B15 D ⁸⁵ Y			<i>P</i> value
		Del/Del (n = 46)	Del/WT + WT/WT (n = 569)	<i>P</i> value	DD (n = 222)	DY (n = 535)	YY (n = 304)	
T (ng/ml)	4.69 ± 1.52	4.76 ± 1.53	4.81 ± 1.46	NS	4.87 ± 1.63	4.72 ± 1.53	4.53 ± 1.38	NS
DHT (ng/ml)	0.31 ± 0.11	0.31 ± 0.12	0.31 ± 0.11	NS	0.31 ± 0.11	0.31 ± 0.12 ^a	0.29 ± 0.10 ^b	0.048
Androsterone-G (ng/ml)	61.2 ± 35.4	61.7 ± 28.8	63.0 ± 35.8	NS	60.4 ± 39.8	63.2 ± 35.1	58.5 ± 32.6	NS
Androstenediol-3G (ng/ml)	1.52 ± 0.92	1.58 ± 0.81	1.58 ± 0.98	NS	1.49 ± 0.86	1.54 ± 0.96	1.51 ± 0.90	NS
Androstenediol-17G (ng/ml)	4.03 ± 2.08	3.28 ± 2.13	4.16 ± 2.09 ^a	<0.001	4.69 ± 2.22 ^a	4.27 ± 2.06 ^a	3.13 ± 1.67 ^b	<0.001
SHBG (nmol/liter)	20.4 ± 7.4	20.4 ± 6.9	20.1 ± 7.0	NS	21.9 ± 7.6 ^{a,b}	20.5 ± 7.6 ^a	19.1 ± 6.7 ^b	<0.001

Values are given as means ± SD. Serum hormone levels were compared by Student's *t* test and *P* values by ANOVA. G, Glucuronide; NS, nonsignificant.

^a $P < 0.05$ vs. Del/Del for UGT2B17 deletion and $P < 0.05$ vs. YY for UGT2B15 D⁸⁵Y.

^b $P < 0.05$ vs. Del/WT for UGT2B17 deletion and $P < 0.05$ vs. DY for UGT2B15 D⁸⁵Y.

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